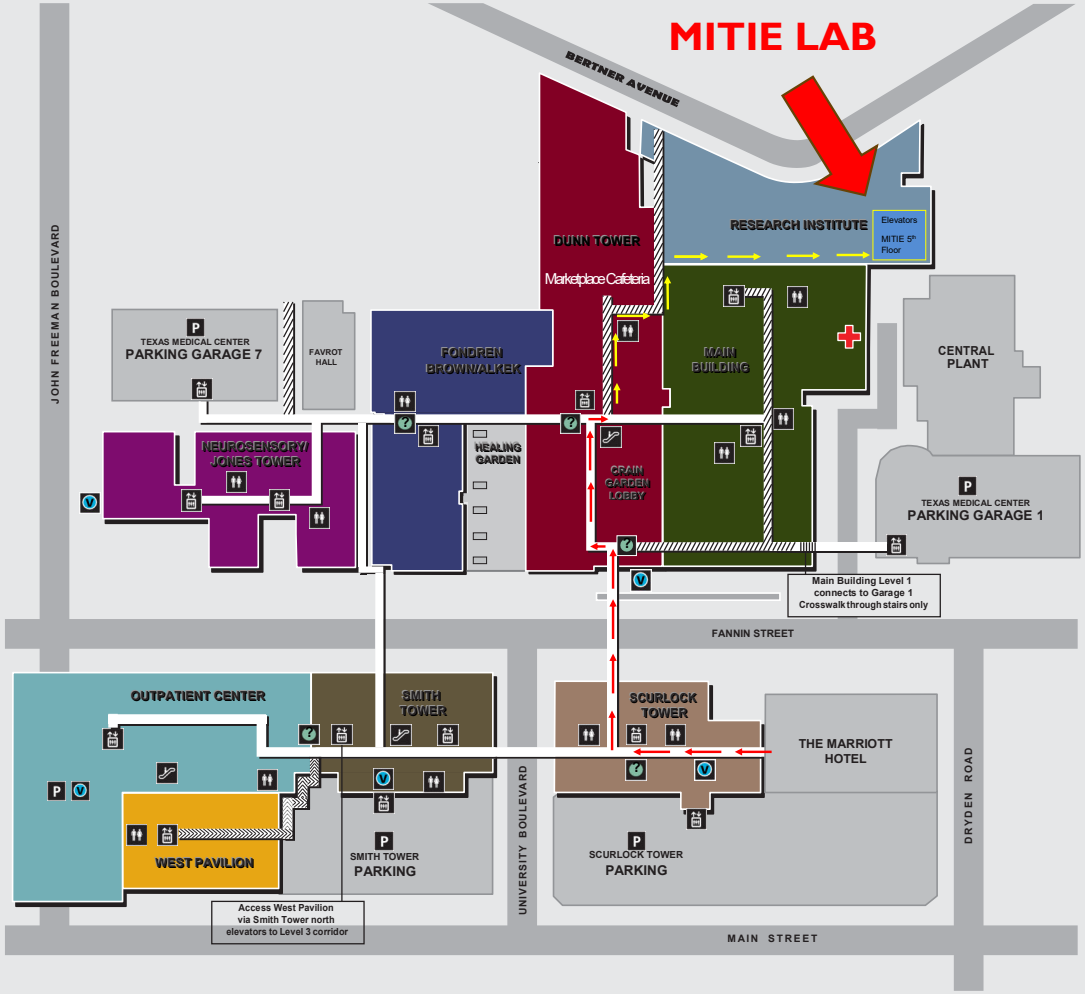
Crosswalk — Level 2

The Level 2 Crosswalk connects most buildings on the campus of The Methodist Hospital. Corridors on Levels 1 and 3 provide access to certain buildings and areas as indicated.

MAP LEGEND

- CROSSWALK
- EMERGENCY ROOM
- ELEVATORS
- ESCALATORS
- INFORMATION
- RESTROOMS
- PARKING
- VALET PARKING



Directions from The Marriott Hotel to MITIE

Follow arrows on map:
Second floor
First floor

- Take the Crosswalk to The Methodist Hospital Scurlock Tower from the 2nd floor of The Marriott
- Continue straight and take the first right to the crosswalk to The Methodist Hospital Dunn Tower
- Head towards the escalators and take them down to the first floor
- Take a right at the bottom of the escalators
- Turn left towards the hallway to the Marketplace Cafeteria
- Make a right at the cafeteria
- Take first left to the Research Institute
- Continue down the hall passed the stairs towards the elevators
- Take elevators up to 5th floor

Methodist The Methodist Hospital

LEADING MEDICINE®

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MITIE LAB LAYOUT

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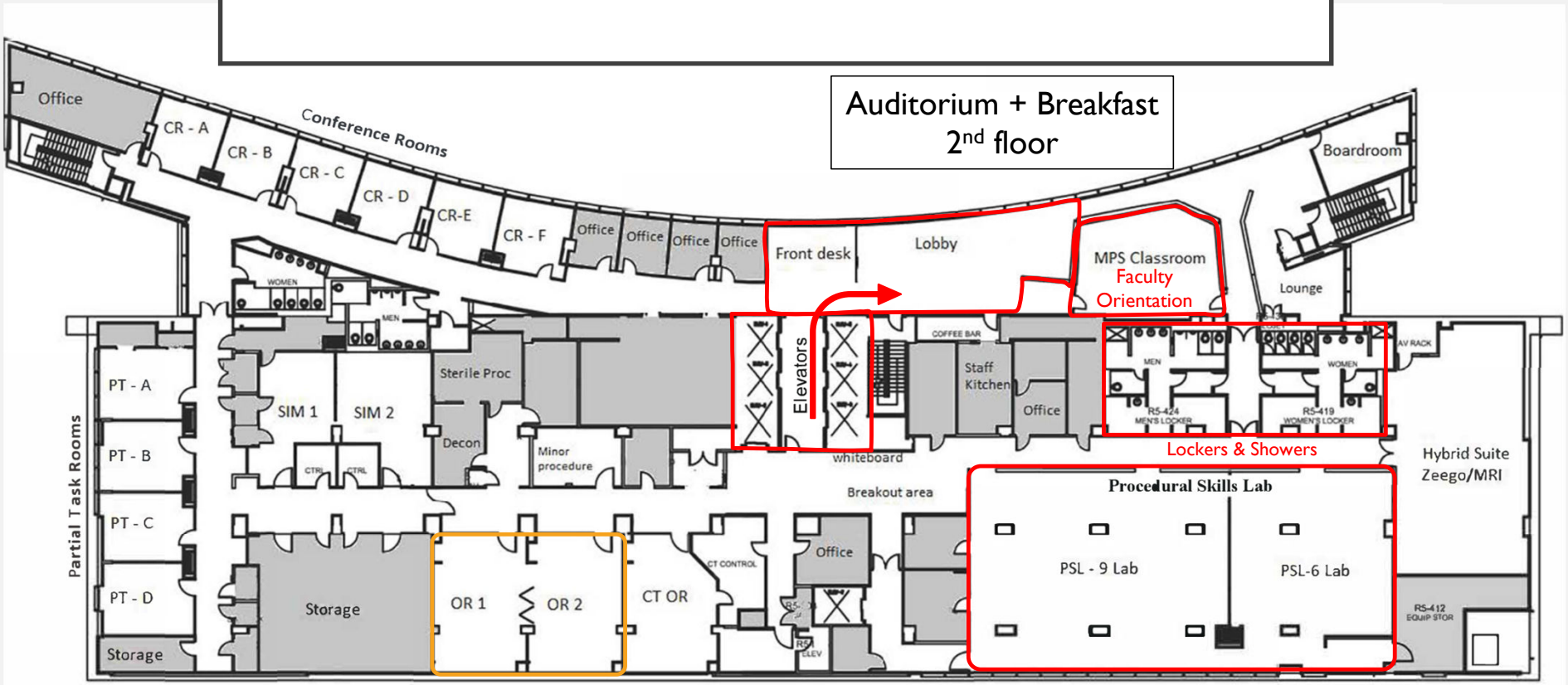


ASOT 2025 WET LAB
RESIDENTS, FELLOWS, AND ATTENDINGS

May 9, 2025

Grayson W Armstrong MD MPH
Chair, ASOT Skills Transfer Lab

Amy Coburn MD
Chair, Annual Meeting

Boonkit Purt MD
Vice President, ASOT

Grant A Justin MD
President, ASOT

ASOT

RESIDENT & ATTENDING WET LAB AGENA

6:30a – 7:00a Registration – *1st fl Research Institute*
Breakfast – *2nd fl Auditorium*

7:00a – 7:30a *Attendee Welcome & Orientation – 2nd fl Auditorium – Barton Blackorby & Boonkit Purt*
Faculty Orientation – 5th fl MPS Classroom– Grayson Armstrong MD

WET LAB	**Orbit & Adnexa**	**Open Globe**	**Anterior Vit / IOL**	**Choose Your Own Adventure**
7:40a – 8:50a	Group A	Group B	Group C	Group D
8:55a – 10:05a*	Group B	Group A	Group D	Group C
10:10a – 11:20p	Group C	Group D	Group A	Group B
11:25p – 12:35p	Group D	Group C	Group B	Group A

**Note: Second Faculty Orientation 9:30-10:00a – 5th fl MPS Classroom – Grayson Armstrong MD*

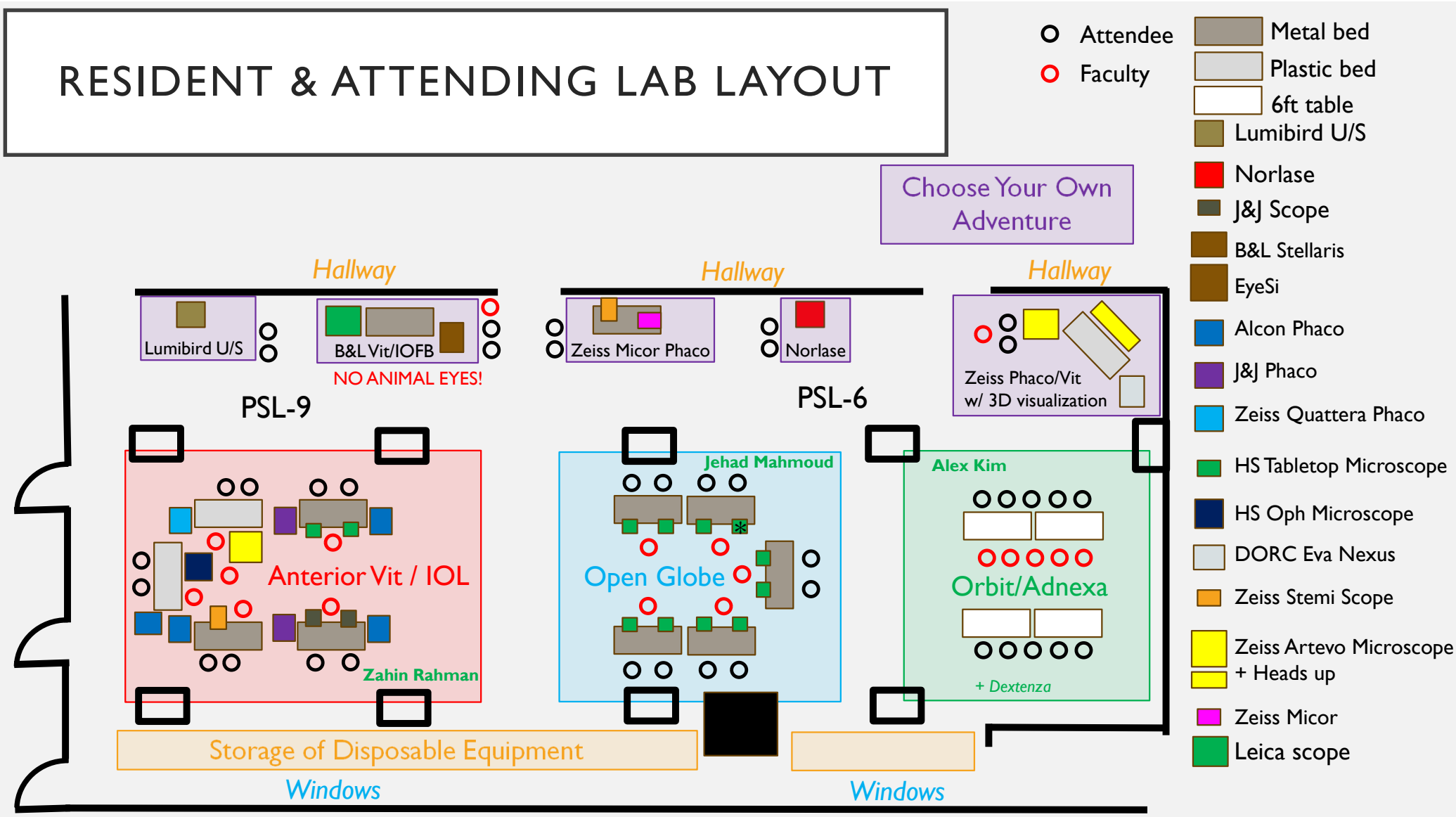
RESIDENT & ATTENDING LAB FACULTY ASSIGNMENTS EARLY SHIFT – 7:40A-10:05A

- Orbit / Adnexa
  - Max Scoville MD
  - Gangadhara Sudar DO FRCSEd FAMS
  - Yunia Irawati MD
  - James Zhao MD
  - Imtiaz Chadry MD
  - Steven Gayer MD (anesthesia block demo)
- Open Globe
  - Marta Stevanovic MD
  - James Auran MD
  - Carlos Andres Wong Morales MD
  - Rupesh Agarwal MD
  - Richard J Blanch FRCOphth
- Anterior Vit & Secondary IOL
  - Yewlin Chee MD
  - Kristen Hawthorne MD
  - Marshall Bowes Hamill MD
  - Grant Justin MD
  - Boonkit Purt MD
  - Sushank Bhalerao MD
  - Lloyd Williams MD
- Choose your Own Adventure
  - Vitrectomy
    - Guy Hunter
    - Sandra Montezuma MD
    - Hossain Nazarai MD
    - Vadivelu Jaya Prakash MD (AIIMS)DNB FICO FAICO

RESIDENT & ATTENDING LAB FACULTY ASSIGNMENTS LATE SHIFT – 10:10A-12:35P

- Orbit / Adnexa
  - Ore-Ofe Adesina MD
  - Charles Soparker MD
  - Ying Chen MD
  - Oded Sagiv MD MHA
  - Amina Malik MD
  - Steven Gayer MD (anesthesia block demo)
- Open Globe
  - Yuntao Hu MD PhD
  - Ferenc Kuhn MD
  - Jose Dalma-Weiszhausz MD
  - Samantha Rodgers MD LTC(P) USA
  - Joanna Queen MD
- Anterior Vit & Secondary IOL
  - Barton Blackorby MD LTC US
  - Christopher Rapuano MD
  - Fasika Woreta MD
  - Masih Uddin Ahmed MD
  - Shehzad Batliwala DO MGM
  - Garvin Davis MD
  - Kanwal Matharu MD
- Choose your Own Adventure
  - Vitrectomy
    - Touka Banaee MD
    - Thamolwan Surakiatchanukul MD
    - S. Natarajan MD

RESIDENT & ATTENDING LAB LAYOUT



ORBIT / ADNEXA SUPPLIES

<u>Station 1</u>: Orbit and Adnexal Trauma

- Educational Objectives:
  - Evaluate and repair marginal eyelid lacerations, treat orbital compartment syndrome, and canalicular lacerations.
  - Discuss the indications for and perform an enucleation.
  - Discuss the indications of and perform a tarsorrhaphy.
  - Discuss the indications for and perform a canthotomy and cantholysis.
  - Discuss indications and timing for orbital fracture repair.

- Learning Objectives:
  - Exploration to identify structures; full-thickness lid marginal laceration repair; reapproximating tarsal plate; tarsorrhaphy; lateral canthotomy/cantholysis for orbital compartment syndrome; enucleation; extraocular muscle securing and suturing
  - Emphasis on anatomic structures; closure in layers; ideal wound construction

Model Used (total needed)
- Goat head (40)

Reusable Supplies (per station/total needed)
- Barraquer Wire Eyelid Speculum (1/10)
- Castroviejo locking needle holder (1/10)
- Curved Hemostats (2/20)
- Desmarres Retractor (2/20)
- Toothed Forceps (1/10)
- Enucleation (or strab) scissors (1/10)
- Enucleation spoon (1/10)
- Muscle Hook (2/20)
- Blunt Westcott scissors (1/10)
- 27g needle on 3cc syringe with BSS to increase retroorbital pressure (1/10)
- Eyelid pins (T-shaped) (2/20)
- Punctal dilators 00 and 0 sizing (1/10)
- 14 blade (or 11 or 15) (1/10)
- Scalpel handles for blades (1/10)

Single use supplies (total needed)
- 4” x 4" gauze (lots)
- Surgical gloves (lots)
- Surgical aprons (lots)
- 6-0 Monocryl suture (vs Plain Gut) (40)
- 6-0 Prolene with P-3 needle (40)
- 6-0 Vicryl with S-29 needle (40)
- Dextenza (40)

OPEN GLOBE SUPPLIES

Station 2: Corneoscleral Open Globe Repair

- Educational Objectives:
  - Gain knowledge and experience taking a history of and managing repair of eye injuries involving the cornea, limbus and sclera
  - Become competent at repair of open-globe injury and understand common complications of open-globe injury.
  - Manage/repair corneal lacerations, limbal and scleral lacerations, and endophthalmitis via tap & inject

- Learning Objectives:
  - Suture simple and complex corneal, limbal, scleral lacerations; peritomy with relaxing incision; muscle isolation and repair; manage uveal prolapse; tap/culture/inject; discuss timing of potential vitrectomy, removal of intraocular foreign bodies

Model Used (total needed)
- Pig eye (40)

Microscopes (total needed)
- Ophthalmic surgical microscopes (10)

Reusable Supplies (per station/total needed)
- 2.4 keratome (1/10) or other blade to make wound
- Paracentesis blade (1/10)
- 25g irrigating cannula (1/10)
- Fine Needle Holder (1/10)
- Toothed Forceps (1/10)
- Vannas Scissor (1/10)
- Tying Forceps (straight) (1/10)
- Tying Forceps (Curved) (1/10)
- Blunt Westcott Scissors (1/10)
- Push pins (T-shaped) (3/30)
- Foam heads (1/10)

Single use supplies (total needed)
- 4"x4" gauze (lots)
- Surgical gloves (lots)
- Surgical aprons (lots)
- 10-0 nylon (40)
- 9-0 nylon (40)
- 8-0 nylon (40)
- Wek Cell spears (40 packs)
- Viscoelastic (80 up from 40)
- 27g needle (40)
- 3cc syringe (40)
- 30g needle (40)
- 1cc syringe (40)
- BSS (120 up from 40)

ANTERIOR VIT / IOL SUPPLIES

<u>Station 3:</u> Vitreous and Secondary IOLs

- Educational Objectives:
  - Gain knowledge and experience taking a history of eye injuries involving complex crystalline lens trauma scenarios with emphasis on capsular integrity, zonular stability, vitreous prolapse and iris injury.
  - Gain proficiency in understanding and practicing lensectomy and be introduced to various intraocular lens implantation techniques in a traumatized eye

- Learning Objectives:
  - Lensectomy; anterior vitrectomy; triamcinolone/kenalog staining; iris management; sulcus IOL; ACIOL, IOL fixation without capsular support; discuss pediatric and adult traumatic cataract and lens capsular violation management in the setting of trauma

Model Used (total needed)
- SimulEYE Complex Phaco model (40)

Microscopes (total needed)
- Ophthalmic surgical microscopes (7)
- Phaco machines (7)

Reusable supplies (per station/total needed)
- Anterior Vit Setup (1/7)
- 2.4 keratome (1/7)
- Paracentesis blade (1/7)
- Toothed forceps (1/7)
- Fine needle holder (1/7)
- Straight tier (1/7)
- Curved Tier (1/7)
- Vannas scissor (1/7)
- Kenalog (1/7)
- Low temp cautery (1/7)
- MST intraocular forceps (1/7)
- MST intraocular IOL cutting scissors (1/7)
- Iris hooks (1 set/7 sets)
- 25g Maxgrip forceps (1/7)
- MVR Blade (1/7)

Single use supplies (total needed)
- Surgical gloves (lots)
- Surgical aprons (lots)
- 30g bent cannula (40)
- 1cc syringe (40)
- 27g needle (40)
- 30g TSK needles (80)
- 10-0 nylon (40)
- 8-0 Prolene (40)
- CV-8 Goretex suture (40)
- Wek Cell spears (40 packs)
- Viscoelastic (40)
- Sheets glide (40)

IOLs (total needed)
- CT Lucia 602 (40)
- MA60AC (40)
- enVista (40)
- ACIOL (40)

CHOOSE YOUR OWN ADVENTURE SUPPLIES

<u>Station 4</u>: Diagnostics & Surgical Equipment

- Educational Objectives:
  - Become familiar with using surgical equipment to plan and execute proper ocular trauma repair

- Learning Objectives:
  - Personalization and customization of: surgical microscopes, heads up displays, phacoemulsification machines, vitrectomy machines, surgical equipment, suture material, intraocular lenses

Model Used
- Pig eyes
- Kitaro kit
- Simuleye tumor model

Reusable supplies (per station/total needed)
- 2.4 keratome (1/3)
- Paracentesis blade (1/3)
- Cystotome (1/3)
- Capsulorhexis foreceps (1/3)
- Curved tier (1/3)
- Straight tier (1/3)
- Toothed forceps (1/3)
- Fine needle holder (1/3)
- Vannas scissor (1/3)
- Vitrectomy setup
- Vitrectomy handpieces
- MVR blade (1/3)

Major equipment (total needed)
- Micor Phaco (1)
- Phaco/Vit machine (2)
- Ophthalmic surgical microscope (3)
- 3D display (1)
- Posterior visualization (2)
- Norlase laser (1)
- Ultrasound (1)

Single use supplies (total needed)
- Malyugin rings
- Surgical gloves (lots)
- Surgical aprons (lots)
- 30g bent cannula (40)
- Cystotome (40)
- Hydrodissection cannula (40)
- 3cc syringe (40)
- 10-0 nylon (40)
- Viscoelastic (40)
- BSS (40)

ASOT 2025 WET LAB
MEDICAL STUDENTS

May 9, 2025

Jared Tuttle BS
Board Member, ASOT

Grayson W Armstrong MD MPH
Chair, ASOT Skills Transfer Lab

Amy Coburn MD
Chair, Annual Meeting

Boonkit Purt MD
Vice President, ASOT

Grant A Justin MD
President, ASOT

ASOT

STUDENT LAB AGENA

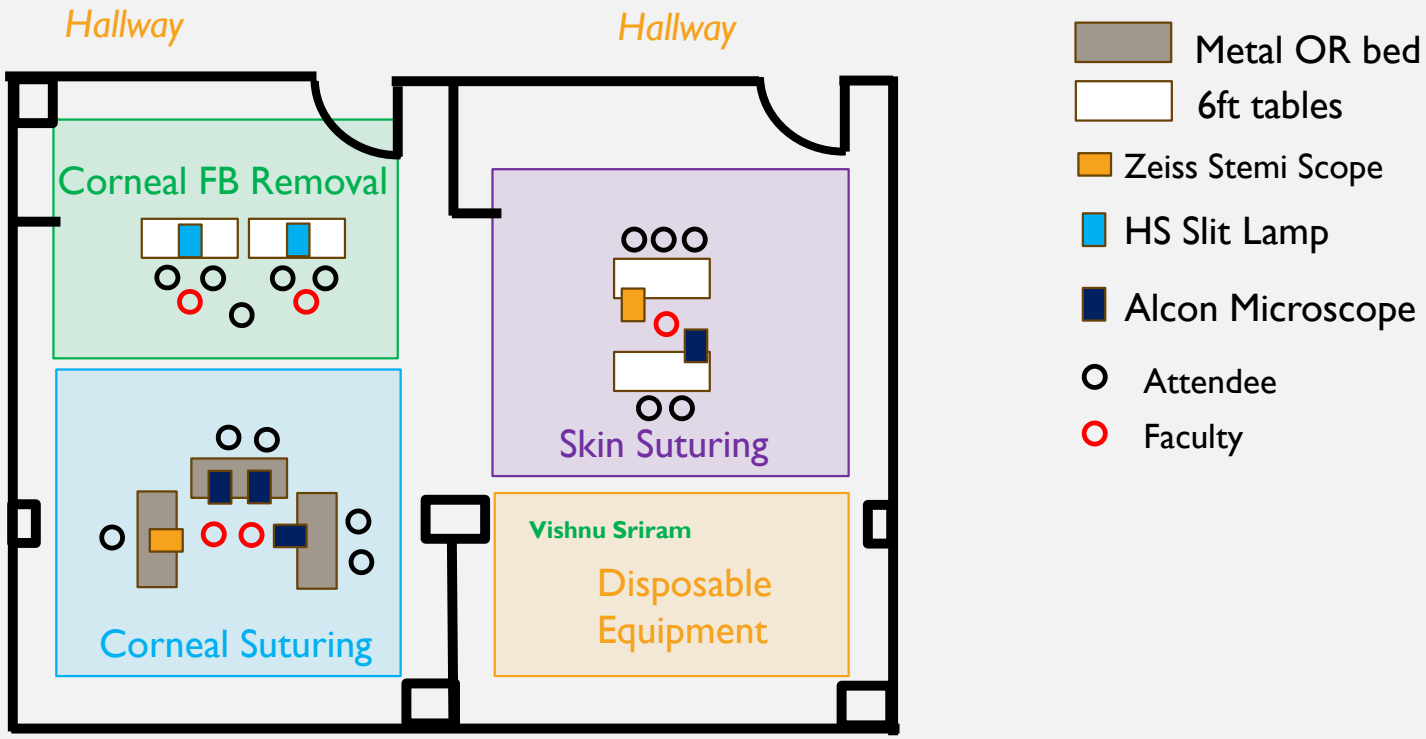
7:30a – 8:00a	Registration – *1st fl Research Institute* Breakfast – *2nd fl Auditorium*		
8:00a – 8:30a	Attendee Orientation, Trauma Lecture – *2nd fl Auditorium (Jared Tuttle, Ankoor Shah)* Faculty Orientation – *5th fl MPS Classroom (Grayson Armstrong)*		
WET LAB	**Corneal Suturing**	**Skin Suturing**	**Corneal FB Removal**
8:40a – 9:10a	Group A	Group B	Group C
9:10a – 9:40a	Group B	Group C	Group A
9:40a – 10:10a	Group C	Group A	Group B
10:10a– 10:20a	**---INTERMISSION---** *Student Trauma Lecture – 5th fl MPS Classroom – Ankoor Shah MD* *Later Faculty Orientation (10:00-10:20a) – 2nd fl Auditorium – Grayson Armstrong MD*		
10:20a – 10:50a	Group D	Group E	Group F
10:50a – 11:20a	Group E	Group F	Group D
11:20a – 11:50a	Group F	Group D	Group E

STUDENT LAB FACULTY ASSIGNMENTS

- 8:00am – 10:10am
  - Ankoor Shah MD PhD – Corneal FB Removal
    - Annette Hoskin PhD MBA – BETTS and Traumatology Education
  - Mona Hussein MSc FRCS (UK) – Corneal suturing
  - Michael Repka MD MBA – Corneal suturing
  - Parag Gandhi MD – Skin Suturing
- 10:40am – 11:50am
  - Donny W Suh MD MBA FACS FAAP – Corneal FB Removal
    - Annette Hoskin PhD MBA – BETTS and Traumatology Education
  - Kalaivani Mohan, MD – Corneal suturing
  - Jennifer Yu MD PhD – Corneal suturing
  - Marguerite Weinert MD – Skin Suturing

STUDENT LAB LAYOUT

OR 1 & OR2



MEDICAL STUDENT WET LAB – CORNEAL SUTURING

<u>Station 5</u>: Basic intraocular skills

- Educational Objectives:
  - Gain experience with microsurgical techniques including corneal suturing
  - Become familiar with the use of surgical microscopes for use in ophthalmic surgery
  - Demonstrate proper handling of surgical instruments used for basic corneal suturing techniques (needle driver, forceps, and scissors) with correct hand positioning and control.
  - Perform at least one basic suturing technique (simple interrupted) on the pig eye with appropriate tension and spacing between stitches.
  - Recognize and correct common errors in suturing technique, such as improper knot tying, uneven tension, or inaccurate needle placement.

- Learning Objectives:
  - Suturing of cornea and ocular tissues; visualization of microsurgical field and tissues using ophthalmic microscopes

Model Used (total needed)
- Pig eyes (30)

Microscopes (total needed)
- Ophthalmic surgical microscopes (or tabletop if necessary) (5)

Reusable supplies (per station/total needed)
- 11 blade (1/5)
- Paracentesis blade (1/5)
- Curved tier (1/5)
- Straight tier (1/5)
- Toothed forceps (1/5)
- Fine needle holder (1/5)
- Vannas scissor (1/5)

Single use supplies:
- 9-0 nylon or 10-0 nylon (30)
- Viscoelastic (60) or KY jelly
- BSS (30) or normal saline
- Weck-cel sponges

MEDICAL STUDENT WET LAB – SKIN SUTURING

<u>Station 6</u>: Basic suturing & surgical microscope management

- Educational Objectives:
  - Gain experience with microsurgical techniques including skin suturing
  - Demonstrate proper handling of surgical instruments used for basic suturing techniques (needle driver, forceps, and scissors) with correct hand positioning and control.
  - Perform at least one basic suturing technique (simple interrupted) on a simulation model with appropriate tension and spacing between stitches.
  - Recognize and correct common errors in suturing technique, such as improper knot tying, uneven tension, or inaccurate needle placement.

- Learning Objectives:
  - Suturing of skin, soft tissues, and ocular tissues

Model Used (per station/total needed)
- Fake skin for suturing (1/5)

Microscopes
- Surgical microscopes (2)

Reusable supplies (per station/total needed)
- Curved tier (1/5)
- Straight tier (1/5)
- Toothed forceps (1/5)
- Fine needle holder (1/5)
- Vannas scissor (1/5)

Single use supplies:
- 6-0 vicryl (30)
- Orange (cut in half) (15)

MEDICAL STUDENT WET LAB – CORNEAL FB REMOVAL

<u>Station 7</u>: Basic suturing & surgical microscope management

- Educational Objectives:
  - Gain an understanding of corneal foreign body removals and approaches for removal
  - Discuss risks of corneal foreign body removal and the potential for a corneal laceration or intraocular foreign body, and the role of seidel testing to assess globe integrity
  - Practice corneal foreign body removal, burring of residual rust, and seidel testing

- Learning Objectives:
  - Corneal foreign body removal, corneal burr for rust removal, and seidel testing

Model Used (total needed)
- Pig eyes (30)

Microscopes
- Slit lamp (2) – Haag Streit + Zeiss

Reusable supplies (per station/total needed)
- Styrofoam head (1/5)
- Wet lab pig eye pins (4/20)
- Corneal burr (1/2)
- 14 blade scalpel (1/5)

Single use supplies:
- 30g needle (30)
- Small metallic FB (30)
- Fluorescein strips (30)